

## Research Article

# Synthesis of Complexes of Azole Derivatives with Ethylenediamine and Biological Evaluation

Hadjer Far<sup>1\*</sup>, Tahar Benaissa<sup>1</sup>, Sofiane Daoudi<sup>1</sup> and Djallal Eddine Adli<sup>2</sup>

<sup>1</sup>Physical Chemistry Studies Laboratory, University, Dr. Moulay Tahar, Saïda – 20000, Algeria

<sup>2</sup>Department of Biology, Faculty of Sciences and Technology, University of Dr. MoulayTahar, Saïda 20000, Algeria

## Abstract

Starting from lauric acid two novel metal complexes of Cu (II) and Co (II), containing azoles derivative nucleus was synthesized by involving multiple-step procedure. This procedure involves four reactions. The first one relates to the esterification of lauric acid using ethanol in the presence of sulphuric acid. As for the second reaction, it is the addition of hydrazine. The third one consists on the cyclization in basic medium, then finally, the coordination with the éthanediamine. In the first part, the resultant compound was characterized by IR and NMR spectroscopy. Its bactericidal activity was evaluated, in the second part, by determining minimum inhibitory concentration (MIC) values and inhibitory zone diameter against gram positive bacteria (*Bacillus subtilis* (BS) and *Listeria monocitgenes* (LM)) and gram negative bacteria such as *Klebsiella pneumonia* (KP) and *Pseudomonasaeruginosa* (PA).

The antifungal activities were tested against four phytopathogenic ungal strains namely *Fusariumgraminearum*, *Aspergillusochraceus*, *Aspergillusparasiticus* and *Penicillium expansum*. Some of the tested compounds displayed promising antibacterial and antifungal activities.

**Keywords:** azoles derivative, complexes, antifungal activities, lauric acid, antimicrobial acrivites

## \*Correspondence

Author: Hadjer Far

Email: hadjer.far@gmail.com

## Introduction

The Carbon-nitrogen bonds are ubiquitous in natural medicines [1]. and organic materials such as azoles [2]. The formation of bonds of this type is among the most important chemical transformations in heterocycle synthesis and medicinal chemistry. Heterocycles are a structural pattern that interning in a multitude of bioactive important natural products [3], pharmaceutical [4], given their wide field of application, as analgesic [5], antidiabetic [6], antimicrobial [7], antiallergic [8-9], anticonvulsant [10], antifungal [11], and finally antidepressants [12], that is why we are interested in the synthesis of azole derivatives, by developing a method of forming carbon-nitrogen bonds, by exploiting the reactivity of the acid function, and who has us among syntheses a derivative from fatty acids by involving reactions.

Esterification, addition of hydrazine and finally cyclization in basic medium. The synthesized azole derivatives have been characterized by different spectroscopic methods, IR, <sup>1</sup>H and <sup>13</sup>C NMR. The latter are merged by coordinate with the éthanediamine [13], leading to schiff-base ligands; the nitrogen site has been used to study their complexation with metallic cations such as Cu<sup>2+</sup> and Co<sup>2+</sup>. The complexes obtained, after spectroscopic characterization by different methods, have been biologically tested in relation to certain molds belonging to the genera: *Aspergillus*, *Alternaria*, *Penicillium* and *Rhizopus*, and on bacteria of the genus: *Staphylococcus*, *listeria*, *Pseudomonas*, *Bacillus* and *Eschirichia*, as well as yeasts of the genus *Candida*.

## Materials and Reagents

All reactions were monitored by thin layer chromatography (TLC) using silica gel F254 supplied by MERCK, using mixture of different polar and nonpolar solvents in varying proportions and spots were observed using iodine as visualizing agent.

All Melting points were determined in open capillary tubes on a BÜCHI 540 melting point apparatus and are uncorrected.

The Infrared spectra of reactants and product in the range of 4000-400 cm<sup>-1</sup> were recorded as potassium bromide discs on a Shimadzu FTIR-8300 Fourier Transform infrared spectrophotometer.

The  $^1\text{H}$  and  $^{13}\text{C}$  NMR Spectra were measured in Chloroform- $d$  ( $\text{CDCl}_3$ ) on Bruker AM 300 MHz Spectrometer (University of Oran, Essenia), relative to the internal standard tetramethylsilane (TMS), and chemical shift values are expressed in parts per million ( $\delta$ , ppm).

#### ***General procedure for the preparation of ethyl laurate B***

This ester was prepared following the standard procedure reported in the literature [14]. Lauric acid (5g, 0.025mol) was dissolved in excess of ethanol (200 mL) with 5 mL of concentrated sulfuric acid and the mixture was refluxed at  $80^\circ\text{C}$  in an oil bath for 6-7 h, the progress of the reaction was monitored by TLC. The excess of acid was neutralized with sodium bicarbonate then the solvent was evaporated and the product was collected.

#### ***General procedure for the preparation of N'-dodecanoylmethanedihydrazide C***

To a solution of lauric ethyl ester B (4.47 g, 0.019mol) in ethanol (100 mL), Nalco (1.76g, 0.019mol) was added and heated for 10 h on oil bath. The progress of the reaction was monitored by TLC.

#### ***General procedure for the preparation of 4-amino-5-undecyl-2,4-dihydro-3H-1,2,4-triazol-3-one D***

A mixture of N'-dodecanoylmethanedihydrazide C, potassium hydroxide (KOH) was refluxed on a oil-bath for 5-7 h. The progress of the reaction was monitored by TLC. After cooling to room temperature, the resulting precipitate was washed with a mixture of water and dichloromethane, and then dried.

#### ***General procedure for the preparation of ligand E***

A mixture of compound (D) (3.82 g, 0.015 mol), ethane-1, 2-diamine (0.45 g, 0.015mol) and ethanol (10 ml) was refluxed on a oil-bath for 6-8h. The progress of the reaction was monitored by TLC. After cooling in an ice bath, the resulting precipitate was filtered, washed with water and dried.

#### ***General procedure for the preparation complexed ligand Co (F)***

To a methanolic solution (1.3g, 0.0024 mol in 20 mL) of the required metal was added a methanolic solution (0.43g, 0.0024mol in 20 mL) of ligand HL in a 1:1 (metal: ligand) molar ratio for Co(II) metal, Then the mixture was gently heated under reflux for 30 minutes and a crystalline colored precipitate was formed at room temperature. The precipitates were filtered out, washed with cold diethyl ether and dried in desiccators.

#### ***General procedure for the preparation complexed ligand Cu (G)***

To a methanolic solution (1.6g, 0.003mol in 20 mL) of the required metal was added a methanolic solution (0.56g, 0.003mol in 20 mL) of ligand HL in a 1:1 (metal: ligand) molar ratio for Cu(II) metal, Then the mixture was gently heated under reflux for 30 minutes and a crystalline colored precipitate was formed at room temperature. The precipitates were filtered out, washed with cold diethyl ether and dried in desiccators

### **Biology**

#### ***Determination of minimum inhibitory concentration (MIC)***

A disk diffusion assay according to the standard protocols (CLSI, 2006) [15]. The solutions of each tested compounds were prepared in DMSO to get a concentration of 100 mg/mL. From this stock solution, serial dilutions of the compounds (100, 50, 25, 12.5, 6.25, 3.125, 1.560, 0.781, 0.390, and 0.195 mg/mL) were prepared. The MIC was recorded in each case as the minimum concentration of the compound, which inhibited the visible growth of the tested microorganism, and DMSO was used as a negative control.

#### ***Antibacterial activity***

In order to evaluate the antibacterial activity four bacteria have been used: *Klebsiella pneumonia* (KP) and *Pseudomonasaeruginosa* (PA) ATCC 70603 and ATCC 27853, as gram negative bacteria and *Bacillus subtilis* (BS) and *Listeria monocitgenes* (LM) as gram positive bacteria.

All microorganisms were obtained from Microbiology Laboratory, Institute of Biology, University of Dr. Moulay Tahar, Saïda. The bacterial strains were maintained on Muller-Hinton agar and the tested compounds were dissolved

in DMSO to make a stock solution of 100 mg/mL and the other concentrations were prepared by dilution. A suspension of the organisms was introduced onto the surface of sterile agar plates, and then incubated at 37 °C for 24 hrs. After incubation, MIC was determined.

### Antifungal activity

The antifungal activities of the synthesized compounds D, E, F and G were studied in different concentrations (1.25, 2.5, 5 and 7.5 µg/ml) against four phytopathogenic fungal strains namely *Fusariumgraminearum*, *Aspergillusochraceus*, *Aspergillusparasiticus*, *Penicilliumexpansum*, The antifungal activity was determined by the Fandohan method [16].

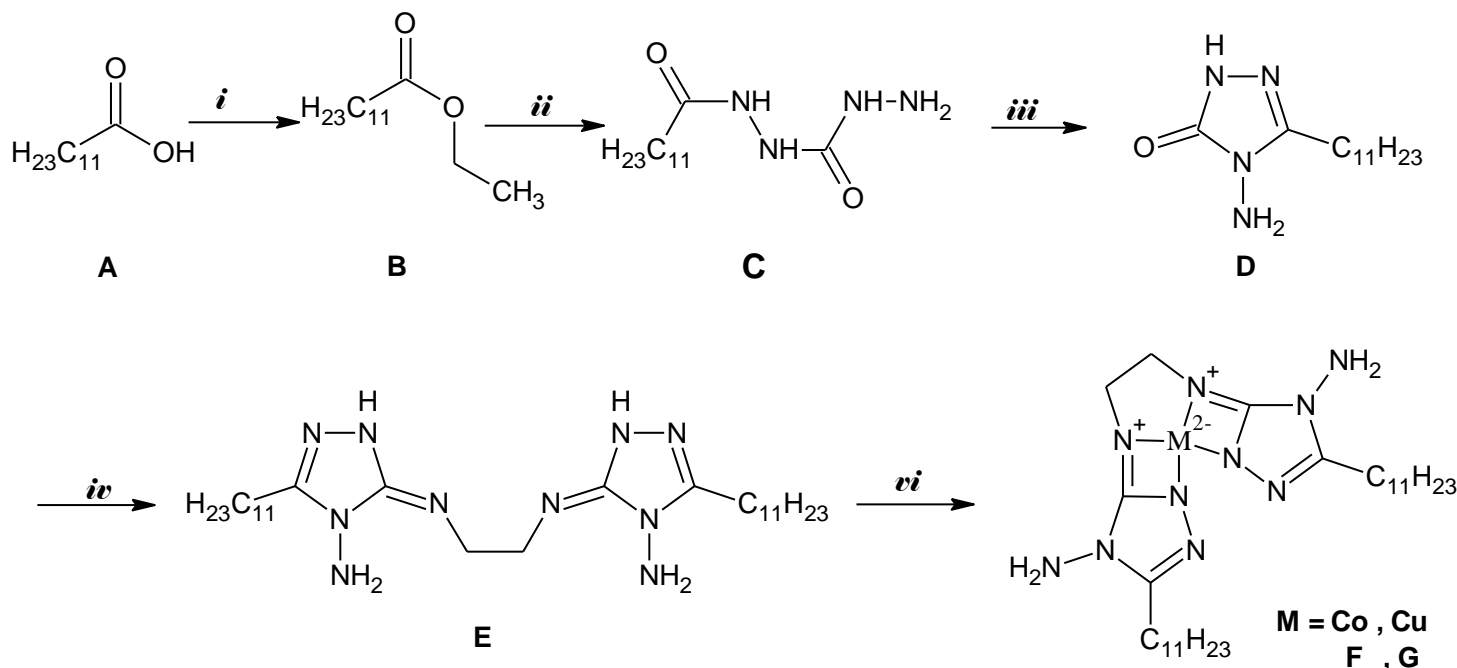
In vitro Screening of Antifungal Activity of synthesized compounds was determined on the potato dextrose agar (PDA) as the growth medium for the tested fungi, and PDA was prepared by dissolving potato extract (200 g), D-glucose (20 g) and agar (15 g) in distilled water (1000 ml). Finally, the medium was transferred to a flask, sealed, sterilized by autoclaving at 121 C for 30 min and cooled down. The compounds were tested at various concentrations and have been incorporated into the PDA culture medium maintained molten at a temperature of 40 to 45°C. After the mixture flow and solidification; Mycelial implants of 6 mm diameter on the pathogen fungi are deposited in the center of the Petri dish containing PDA medium with 04 concentrations for each synthesized compounds to be studied. The Petri-dishes were incubated at 28°C for 3-5 days. Taking into account the mycelial growth of the control [17]. After the completion of incubation period, the Relative inhibition rate of the circle mycelium compared to blank assay was calculated via the following equation:

$$\text{Antifungal index (\%)} = [(1 - Da) / Db] \times 100\% \quad (1)$$

Where: *Da* is the diameter of the test growth area, *Db* is the diameter of the growing area of the control.

### Results and Discussion

The target compounds F and G were synthesized by a multiple-step procedure as shown in **Scheme 1**. The synthetic route started from esterification of lauric acid 1 using ethanol in the presence of sulphuric acid. This ester reacted with nalco in ethanol to give compound C. 4-amino-5-undecyl-2, 4-dihydro-3H-1,2,4-triazol-3-one (D) was obtained by ring closing reaction upon treating compound C with potassium hydroxide in ethanol. This compound was converted to give the ligand LH by ring-alkylation reactions using 4-amino-5-undecyl-2,4-dihydro-3H-1,2,4-triazol-3-one (D) with ethylenediamine (2:1). Finally, the products F and G were obtained by refluxing compound E with the required metal in methanol.



**Reagents and conditions**

(i) C<sub>2</sub>H<sub>5</sub>OH, and conc. H<sub>2</sub>SO<sub>4</sub>, reflux for 8 h; (ii) NH<sub>2</sub>NHCO(NHNH<sub>2</sub>) and C<sub>2</sub>H<sub>5</sub>OH, reflux for 5 h; (iii) KOH and C<sub>2</sub>H<sub>5</sub>OH, Reflux (iv) ethylenediamine and C<sub>2</sub>H<sub>5</sub>OH, (vi) metal (Co, Cu) and CH<sub>3</sub>OH.

Both the free ligand and its metal complexes formations were detected by thin layer chromatography (TLC) via their R<sub>f</sub> which were different from starting materials. The obtained complexes are of various colours varied from brick red to violet colour. It is to note that these colours are different from the ligand colour, which indicate that the colours formed depend on the metal ions. The melting points of the complexes are different (higher) than that of the ligand an evidence for complexation. The physical and analytical data of all the compounds studied has been summarized in **Table 1**.

**Table 1** Physical properties and analytical data of the synthesized compounds

Compound	Colour	Yield, %	Melting point, °C	R <sub>f</sub> value	Solvent system CHCl <sub>3</sub> /CH <sub>3</sub> OH
B		81.5	liquid	0.65	8/2
C		81	Liquid	0.75	8/2
D		92.75	Liquid	0.9	8/2
E	White	60.15	120	0.83	8/2
F	Red	78.5	171-172	0.85	8/2
G	Violet	73.03	208	0.78	8/2

The IR spectra in the (4000–400 cm<sup>-1</sup>) region provide information regarding the coordination mode in the complexes were analyzed by comparison with the data for the free ligand. The IR data of the ligand and complexes are shown in **Table 2**.

**Table 2** The characteristic infrared absorptions in  $\bar{\nu}$  cm<sup>-1</sup> of the synthesized compounds B-G

Comp.	-C-H	C=O	NH <sub>2</sub> /NH	C-O-C	C=N	C-N	M-N
B	2976.0	1739.7	-	1016.4	-	-	-
C	2975.0	1737.7	3483.2	-	-	-	-
D	2925.8	1739.7	3460.1	-	-	1109.0	-
E	2923.9	-	3342.4	-	1566.1	1109.0	-
F	2923.9	-	3134.1-3219.0	-	1585.4	1058.8	1375.2
G	2954.7	-	3163.0-3276.8	-	1606.6	1041.5	1373.2

<sup>1</sup>H NMR (300MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) of **B**: 0.903 (3H, C1), 4.116 (2H, C2), 2.292 (2H, C4), 1.618 (2H, C5), 1.257 (2H, C6, C7, C8, C9, C10, C11, C12, C13), 0.88 (3H, C14). <sup>13</sup>C NMR (300MHz, CDCl<sub>3</sub>),  $\delta$  (ppm): 14.229 (C1, C14), 59.796 (C2), 174.068 (C3), 34.396 (C4), 24.987 (C5), 29.612 (C6, C7, C8, C9, C10, C11), 31.918 (C12), 22.695 (C13).

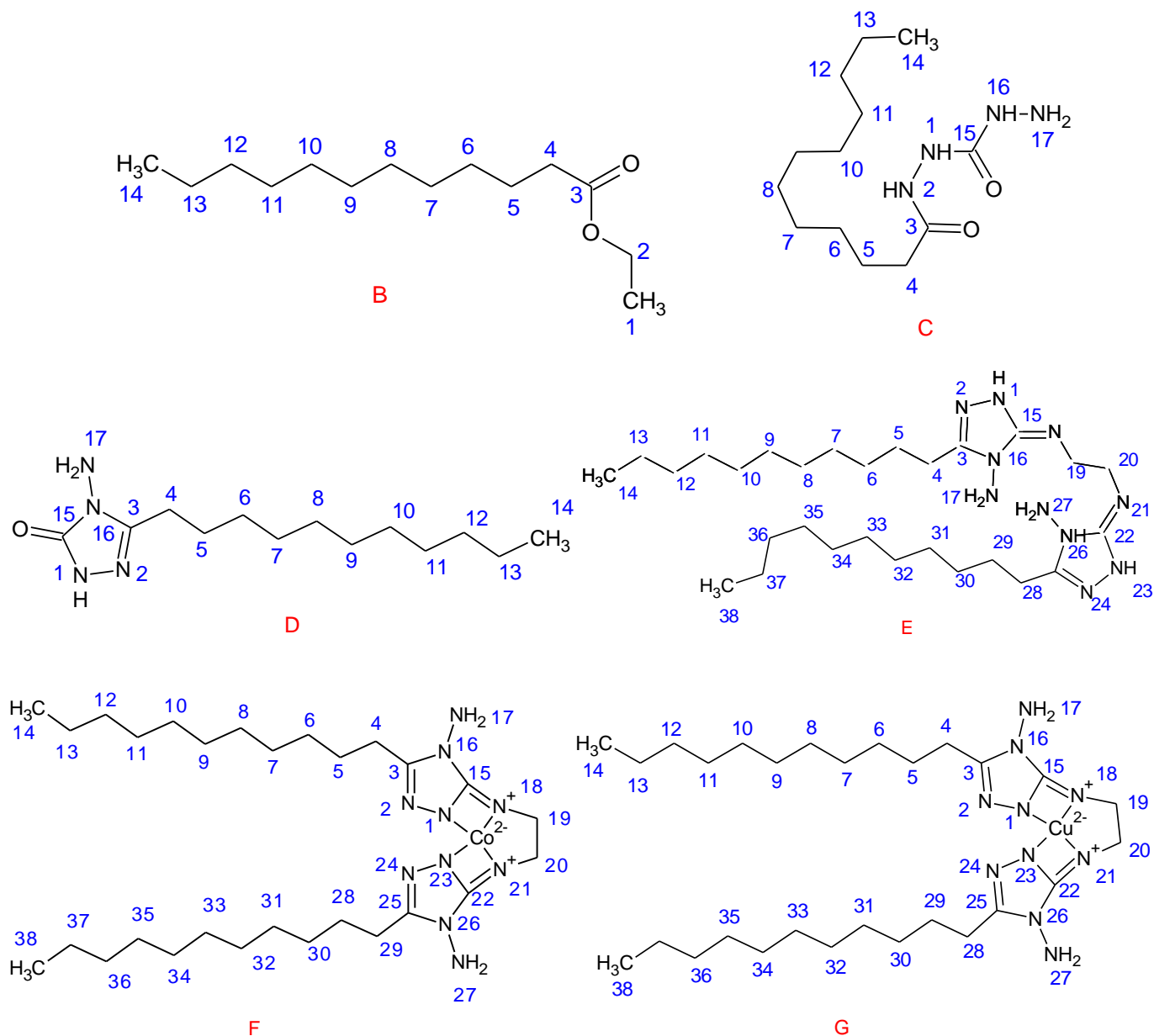
<sup>1</sup>H NMR (300MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) of **C**: 5.026 (H, NH 2, 1, 16), 4.135 (2H, NH<sub>2</sub> 17), 2.285 (2H, C4), 1.253 (2H, C6, C7, C8, C9, C10, C11, C12, C13), 1.624 (2H, C5), 0.854 (3H, C14). <sup>13</sup>C NMR (300MHz, CDCl<sub>3</sub>),  $\delta$  (ppm): 14.249 (C14), 173.703 (C3), 34.394 (C4), 24.730 (C5), 29.453 (C6, C7, C8, C9, C10, C11), 31.605 (C12), 22.464 (C13), 151.767 (C15).

<sup>1</sup>H NMR (300MHz, CDCl<sub>3</sub>),  $\delta$  (ppm) of **D**: 5.075 (H, NH 1), 4.015 (2H, NH<sub>2</sub> 17), 2.191 (2H, C4), 1.044 (2H, C6, C7, C8, C9, C10, C11, C12, C13), 1.522 (2H, C5), 0.786 (3H, C14). <sup>13</sup>C NMR (300MHz, CDCl<sub>3</sub>),  $\delta$  (ppm): 14.171 (C14), 34.394 (C4), 17.341 (C5), 29.358 (C6, C7, C8, C9, C10, C11), 31.814 (C12), 22.748 (C13).

<sup>1</sup>H NMR (300MHz, D<sub>2</sub>O),  $\delta$  (ppm) of **E**: 2.077 (H, NH 1,2,3), 4.698 (2H, NH<sub>2</sub> 17, 27), 2.846 (2H, C4, C28), 1.133 (2H, C6, C7, C8, C9, C10, C11, C12, C13, C37, C36, C35, C34, C33, C32, C31, C30), 1.762 (2H, C5, C29), 1.007 (3H, C14, C38), 2.846 (2H, C19, C20).

<sup>1</sup>H NMR (300MHz, D<sub>2</sub>O),  $\delta$  (ppm) of **F**: 4.697 (2H, NH<sub>2</sub> 17, 27), 2.626 (2H, C4, C38), 1.758 (2H, C6, C7, C8, C9, C10, C11, C12, C13, C37, C36, C35, C34, C33, C32, C31, C30), 2.011 (2H, C5, C29), 1.001 (3H, C14, C38), 2.013 (2H, C19, C20). <sup>13</sup>C NMR (300MHz, D<sub>2</sub>O),  $\delta$  (ppm): 15.842 (C14, C38), 16.043 (C4, C28), 43.815 (C6, C7, C8, C9, C10, C11, C12, C13, C37, C36, C35, C34, C33, C32, C31, C30), 19.176 (C5, C29), 44.267 (C19, C20).

<sup>1</sup>H NMR (300MHz, D<sub>2</sub>O),  $\delta$  (ppm) of **G**: 4.699 (2H, NH<sub>2</sub> 17, 27), 1.910 (2H, C6, C7, C8, C9, C10, C11, C12, C13, C37, C36, C35, C34, C33, C32, C31, C30), 1.911 (2H, C5, C29), 1.026 (3H, C14, C38), 1.910 (2H, C19, C20).



**Figure 1** Schematic drawings developed according to ChemSketch of structures B, C, D, E, F and G for  $^1\text{H}$  and  $^{13}\text{C}$  NMR purposes

### Antibacterial activities tests

The synthesized compounds have been tested in vitro for their antibacterial activity against four bacteria, such as *Klebsiella pneumonia* (KP) and *Pseudomonasaeruginosa* (PA) as gram negative bacteria and *Bacillus subtilis* (BS) and *Listeria monocitgenes* (LM) as gram positive bacteria. The obtained results are summarized in **Table 3**.

**Table 3** Antibacterial activity of the synthesized compounds

Strains	Compound and values of MIC in mg/mL			
	G	F	E	D
PA	0.390	3.125	00.00	12.5
BS	25	50	00.00	25
LM	12.5	25	12.5	12.5
KP	12.5	25	00.00	50

As it is shown in Table 3, the results of evaluating the antibacterial properties suggest that all compounds have very good potential to act as antibacterial agents. The values of MIC ranged between: 0.195-100 mg/mL

**Antifungal activities tests**

The evaluation of antifungal activity revealed that all the synthesized compounds have a significant biological activity, the best activity was observed at a concentration of 7.5 µg/mL against all the tested fungi. The results of in vitro antifungal activities are presented in **Table 4**.

**Table 4** Relative inhibition rate (%) of synthesized compounds against tested fungal strains

		Concentration [µg/ml]	D	E	F	G
Relative inhibition rate (%)	<i>F.graminearum</i>	1.25	53.33	23.33	53.33	28.57
		2.5	60	33.33	56.67	40.48
		5	63.33	40.00	60.00	52.38
		7.5	66.67	50.00	66.67	59.52
	<i>A.parasiticus</i>	1.25	9.20	29.09	26.36	52.73
		2.5	29	43.53	35.25	63.64
		5	35	58	50.00	65
		7.5	36.60	68.35	62.32	70.91
	<i>A.ochracus</i>	1.25	16.67	35.71	40.48	38.55
		2.5	50	52.38	45.24	55.47
		5	54.76	59.52	57.14	62.09
		7.5	71.43	76.19	59.52	79.22
<i>P.expansum</i>	1.25	15.69	27.00	20.00	44	
	2.5	35.08	39.12	33.00	61.22	
	5	42.46	60.16	51.00	73.11	
	7.5	64	80.43	71.20	83.44	

**Conclusion**

In summary, we have described the synthesis of the ligand HL and his metal complexes, IR, NMR spectral techniques were used to confirm their formation. The newly synthesized compounds were evaluated for their antibacterial and antifungal activity. The results of biological tests and the in vitro antifungal activity indicated that most of the synthesized compounds exhibited promising results. These compounds can be considered as lead molecules for future investigations.

**Acknowledgment**

The authors are grateful to Mrs HIDOUR Hanaa (University of Oran, Es-Senia) for recording the NMR spectra.

**References**

- [1] J. Appenzeller, S. Tilvi, M. T. Martin, J. F. Gallard, H. El-Bitar, E. T. H. Dau, & Al-Mourabit, A. Benzocceptins A and B with a unique benzocyclobutane skeleton and nagelamide S and T from Pacific sponges. *Organic letters*, 2009, 11(21), 4874-4877.
- [2] T. B. Nguyen, L. Ermolenko, & A. Al-Mourabit. *Journal of the American Chemical Society*, 2012, 135(1), 118-121.
- [3] Y. S. Prabhakar, V. R. Solomon, M. K. Gupta & S. B. Katti, QSAR studies on thiazolidines: a biologically privileged scaffold. In *QSAR and Molecular Modeling Studies in Heterocyclic Drugs II*. Springer, Berlin, Heidelberg.2006. p. 161-249. Lesyk, R. B., & Zimenkovsky, B. S. (2004). *Current Organic Chemistry*, 8(16), 1547-1578.
- [4] J.W.W. Chang, X. Xu, P.W.H. Chan, *Tetrahedron Lett*, 2007, 48,245-248.
- [5] E. J. Glamkowski, J. M. Fortunato, T. C. Spaulding, J. C. Wilker, & D. B. Ellis, *Journal of medicinal chemistry*, 1985, vol. 28, no 1, p. 66-73
- [6] a) R.B. Chapleo, G.P. Ann. Fagan, *Drug Data Rep.* 1993, 15, 59. b) *Chem. Abstr.* 1992, 117; 90283.
- [7] A. G. Kamat, G. S. Gadaginarnath, *Indian J.Chem*, 1994, Sect. B, 33, 255-259.

- [8] a) P. C. Unangst, D. T. Connor, S. R. Stabler, R. J. Weikert, M. E. Carethers, J. A. Kennedy, ... & M. C. Conroy, Novel indolecarboxamidotetrazoles as potential antiallergy agents. *Journal of medicinal chemistry*, 1989, vol. 32, no 6, p. 1360-1366. b) P. C. Unangst, M.E. Carethers, W. Webster, G.M. Janik, L.J. Robichaud, *J. Med Chem.* 1984, 27, 1692-1633.
- [9] a) L. J. Robichaud, S. F. Stewart, & R. L. Adolphson, CI-922—A novel, potent antiallergic compound—I. Inhibition of mediator release in vitro. *International journal of immunopharmacology*, 1987, vol. 9, no 1, p. 41-49. b) C.D. Wright, M. D. Hoffman, Thueson, D. O. *Luekocyte Biol.* 1987, 42, 30-35.
- [10] J. Hazarika, & J. C. S. Katakya, Studies on biologically active heterocyclics. Part VII synthesis and biological activity of some new 3-substituted 5-(2-chlorophenyl)-1, 3, 4-oxadiazol-2-thiones and their derivatives. *Indian Journal of Heterocyclic Chemistry*, 1998, 7(3), 197-200.
- [11] T. P. Dabhi, V. H. Shah, & A. R. Parikh, *Indian Drugs*, 1992, vol. 54, p. 98-98.
- [12] H. P. Shah, B. R. Shah, J. J. Bhatt, N. C. Desai, P. B. Trivedi, & N. K. Undavia, Synthesis of 2, 5- Disubstituted 1, 3, 4- Oxadiazoles as Potential Antimicrobial, Anticancer and Anti- HIV Agents. *Ind. J. Chem.*, 1998, vol. 29, no 39.
- [13] M. H. Habibi, M. Montazerzohori, K. Barati, R. W. Harrington, W. Clegg, J-H. Choi, *Analytical Sciences. The Japan Society for Analytical Chemistry.* 2007, 23, 117.
- [14] B.S. Furniss, A.J. Hannford, P.W.G. Smith, A.R. Tatchell, *Vogel's Text Book of Practical Organic Chemistry.* John Wiley & Sons: New York, 5th edition, 1989, pp. 1076.
- [15] W. CLSI, *Clinical and laboratory standards institute methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically. Approved Standard M7-A7, CLSI, seventh ed., PA, USA, 2006.*
- [16] P. Fandohan, J. D. Gbenou, B. Gnonlonfin, K. Hell, W. F. Marasas, & M.J. Wingfield, Effect of essential oils on the growth of *Fusarium verticillioides* and fumonisin contamination in corn. *Journal of agricultural and food chemistry*, 2004, 52(22), 6824-6829.
- [17] S. Y. Wang, P. F. Chen, & S. T. Chang, Antifungal activities of essential oils and their constituents from indigenous cinnamon (*Cinnamomum osmophloeum*) leaves against wood decay fungi. *Bioresource technology*, 2005, 96(7), 813-818.

## Publication History

Received 04<sup>th</sup> Aug 2018  
Revised 20<sup>th</sup> Sep 2018  
Accepted 28<sup>th</sup> Sep 2018  
Online 30<sup>th</sup> Oct 2018

© 2018, by the Authors. The articles published from this journal are distributed to the public under “**Creative Commons Attribution License**” (<http://creativecommons.org/licenses/by/3.0/>). Therefore, upon proper citation of the original work, all the articles can be used without any restriction or can be distributed in any medium in any form.